Growth of *Dendranthema × grandiflorum* (Ramat.) Kitamura under Various Spectral Filters

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**Abstract.** ‘Spears’ (nonpinched and pinched) and ‘Yellow Mandalay’ (pinched) chrysanthemums were grown in growth chambers equipped with panels filled with liquids that served as spectral filters. Light quality was altered by reducing blue light, increasing red : far-red (R : FR) light, or reducing R : FR. Control panels did not selectively alter light transmission. Photosynthetic photon flux was the same in all chambers. All plants grown under increased R : FR filters had reduced height, reduced internode length, and increased chlorophyll content compared to controls. Reduction in blue light decreased chlorophyll content of pinched plants compared to controls. Pinched plants grown under increased R : FR light and long days developed fewer nodes than controls due to the formation of abnormal capitula; the controls and plants from the other treatments developed more nodes before producing similarly abnormal capitula. Stem diameter and leaf area did not differ due to treatments.

Chemical growth regulators are often used to maintain desired growth and form of potted chrysanthemums. However, increasing awareness of the impact of chemicals on the environment and strict U.S Environmental Protection Agency regulations may limit or prohibit the use of many of these chemicals. For example, butanedioic acid mono(2,2-dimethylhydrazide) (daminozide, B-Nine, Alar) is frequently used to control potted chrysanthemum height. The use of daminozide on edible crops was recently prohibited.

Light quality has been demonstrated to influence many aspects of plant growth and morphology (Borthwick and Cathey, 1952; Borthwick et al., 1952; Mortensen and Stromme, 1987; Vince-Prue and Canham, 1983). Phytochrome, the primary pigment controlling photomorphogenesis, has two forms, P<sub>r</sub> and P<sub>t</sub>, which have peak absorptions in red (660 nm) and far-red wavelength (730 nm) light, respectively. Upon absorption, a phytochrome chromophore converts to the other form (Borthwick et al., 1952). Light quality determines phytochrome equilibrium (φ) in vivo, which, in turn, determines plant morphology (Holmes and Smith, 1977). Phytochrome equilibrium is often expressed as the ratio of P<sub>r</sub> to total phytochrome (φ = P<sub>r</sub> : P<sub>t</sub>). P<sub>r</sub> : P<sub>t</sub> is determined by integrating the quantitative effects of all wavelengths between 350 and 850 nm on phytochrome equilibrium (Sager et al., 1988).

A plant canopy acts as a selective filter that absorbs red (R) and blue (B) but is relatively transparent to FR (Kasperbauer, 1988). Under a leaf canopy, plants receive light with a decreased R : FR or reduced P<sub>r</sub> : P<sub>t</sub> compared to full sunlight. These plants are generally taller and have a different chloroplast arrangement than plants grown in full sun (Eskins and Duysen, 1984; Kasperbauer, 1988). Plants receiving a relatively low R : FR also exhibit longer internodes, less branching, and larger and thinner leaves (Kasperbauer, 1988).

Since phytochrome absorbs in the B region (400-500 nm) as well, B may also affect plant morphology by influencing phytochrome or another, unidentified photoreceptor (cryptochrome) that affects plant morphology (Duell-Pflaum and Wellmann, 1982). Dritz and Sager (1990) reported that soybeans grown under a blue-deficient, narrow-band light source resembled shaded plants.

Materials that selectively filter out specific radiation wavelengths entering a greenhouse may allow manipulation of plant growth. Mortensen and Stromme (1987) grew several plant species under selective spectral filters. Plants continually grown with increased R : FR were shorter and darker green with more branches, but had smaller leaves and lower dry weights than those grown in reduced R : FR. More information is needed on the effects of altered spectral radiation on the growth and development of floricultural crops. This study was conducted to determine the effects of B<sub>2</sub>, R<sub>2</sub>, and FR-filtered light on two commercial, pinched pot chrysanthemums, ‘Spears’ and ‘Yellow Mandalay’.

**Materials and Methods**

**General procedures.** Ten growth chambers were constructed in house as an attached lean-to on the side of an existing greenhouse. The glass wall on the south side of the greenhouse was removed and vertical aluminum panels were installed 55 cm apart and 2.5 cm from the bottom of the chambers at each glazing bar, forming reflective walls for each chamber. The bottom of the chambers was a continuous aluminum pan that was the width and length of all 10 chambers. A 7-cm-wide strip of flat black paint was applied to the pan centered beneath each vertical wall to prevent light from reflecting between adjacent chambers. The back of the chambers was a sheet of black polyethylene between two layers of white polyethylene that could be rolled up to allow access to the chambers. The front of each growth chamber was a 54 × 140-cm sheet of transparent hollow polycarbonate with cross sections of columns 6 mm deep and 11 mm wide (Polygal, Janesville, Wis.) treated for resistance to ultraviolet (UV) radiation and sealed at one end with caulk ing compound. A fan located at the top of each growth chamber

**Abbreviations:** B, blue; FR, far red; P, phytochrome; PPF, photosynthetic photon flux; R, red; UV, ultraviolet.
circulated air from the attached greenhouse through each unit. The greenhouse air was heated or cooled to modulate temperature in the chambers. Thermistors in each chamber were connected to a computer and provided constant monitoring of chamber temperature. Plants were placed in each chamber on a white plastic grid with 1.75 × 1.75-cm openings (manufactured for use as a diffusing panel for fluorescent light fixtures). The plastic grid was secured 14 cm below the edge of the pan. Pollock et al. (1990) thoroughly describe the chambers used.

The polycarbonate panel channels were filled with solutions chosen for their spectral filtering properties and resistance to photodegradation. The spectral filter solutions were a red dye #259 (CIBA-GEIGY, Greensboro, N.C.) that filtered out much of the B and green light, a blue dye #178 (CIBA-GEIGY) that filtered much R but not FR, and CuSO$_4$·5H$_2$O, which filtered a greater proportion of FR than R light. Two controls were used, water- and air- (normal content of the panels) filled panels. Data from representative spectroradiometer scans of the selective filtering solutions and full sunlight are shown in Fig. 1. Scans of the controls (between 330 and 850 nm) differed from full sunlight only in intensity (data not presented). The percent transmission for each of the selective filters (compared to full sun) is presented in Fig. 2. R : FR ratios (narrow and broad band) and P$_{fr}$ : P$_{int}$ determined from the formulas of Sager et al. (1988), varied in a similar manner for all treatments (as R : FR increased, P$_{fr}$ : P$_{int}$ increased) (Table 1); therefore, description of the light environment for most of the remainder of this paper will be as R : FR because this measurement is more easily obtainable for researchers who do not have access to a spectroradiometer.

For most experiments, the 10 chambers allowed two replications of each of five treatments. For the remaining experiments, eight chambers were used and only four treatments were evaluated (two replications of each treatment). All experiments were repeated.

All concentrations of the spectral filtering solutions were adjusted to provide similar photosynthetic photon flux (PPF) transmission, i.e., a 40% to 45% reduction in PPF through the panels. The CuSO$_4$·5H$_2$O solution (16%, w/v) was the PPF reference for all other treatment concentrations. A gray, neutral-density shade material was placed over the control panels to provide an equal reduction in PPF. All light measurements were made with an LI-1800 spectroradiometer equipped with a LI-1800-10 remote cosine sensor (LI-COR, Lincoln, Neb.). Light measurements were taken at about solar noon on cloudless days. Measurements were expressed as percentage of full sun. PPF measurements were also taken with a LI-COR LI-185-B quantum sensor. Quantum sensor readings were slightly higher because the cut-off at 400 and 700 nm is not as sharp as with the spectroradiometer; however, percent transmission was unchanged. Although PPF is a single measurement over a wide (300 nm) band width and was equal for all treatments, the wavelengths contributing to PPF varied greatly among selective filters (Fig. 1). The differences resulted in different levels of photosynthetically active radiation among chambers (McCree, 1972).

Influence of spectral filters on nonpinched 'Spears' chrysanthemums (Expt. 1). Five adjacent growth chambers were one replication (block) and the next five were the second replication (block) with the treatments randomly assigned to each replication. Spectral qualities of the panels were checked weekly. The red and blue dye solutions were replaced every 2 weeks because preliminary experiments indicated that red dye had changed spectrally after 3 weeks and the blue dye after 4 weeks. Copper sulfate was changed at the end of each experiment.

Forty uniform, rooted 'Spears' cuttings (Yoder Bros., Pendleton, S.C.) were planted individually in 11.25-cm (550 cm$^3$) square, opaque, green geranium plastic pots (Spartan Plastics,
Spartanburg, S.C.) containing commercial potting media (Fafard #3B, Fafard, Anderson, S.C.). Four pots were placed in each chamber on 27 Apr. 1989. The plants were fertilized every other day with 300N–132P–249K (mg liter⁻¹) (from Peter’s 20-20-20; Peter’s, Fogelsville, Pa.) for the first week and then with a solution of 0.0026 N KNO₃ and 0.0073 N Ca(NO₃)₂ until the experiment was terminated. Water was applied as needed between fertilizations. All pots received the same fertilizer/water regime. Temperatures in the chambers ranged from a night average of 18°C (SE ± 0.1) to a day average of 28°C (SE ± 0.4); the 24-h mean was 24°C (SE ± 0.3). Light intensity in the chambers was 55% to 60% of natural light (natural light is ~1700 µmol m⁻² s⁻¹ PPF on a clear day at solar noon in South Carolina). Plants were not pinched. The experiment was terminated on 10 May. Data were collected on plant height above pot rim, chlorophyll content, number of leaves, stem diameter, internode length, and leaf area. Chlorophyll was extracted by the method of Moran and Porath (1980). Five leaf disks (0.28 cm²/disk) from each plant (one disk from each of the most recently expanded leaves) were removed, chlorophyll was extracted in N,N-dimethylformamide for 48 to 72 h in darkness at 4°C (five disks/7 ml), and absorbance measurements were made with a Spectronic 1001 (Bausch and Lomb, Rochester, N.Y.). Chlorophyll content was calculated by the formulas of Moran (1982). Stem diameter was measured between the seventh and eighth nodes. Because compact internode length of CuSO₄-filter-grown plants were difficult to measure, internode length was measured as the average length between the seventh and ninth nodes. Leaf area was measured using a LI-COR Leaf Area Meter Model 3100. Polycarbonate panels were cleaned, randomly filled with solutions or air, replaced randomly as described previously, and the experiment was repeated on 18 May 1989, terminating 31 May.

**Influence of spectral filters on pinched ‘Spears’ chrysanthemums** (Expt. 2). Experimental conditions were the same as Expt. 1, except plants were placed in the chambers on 15 May 1989 and pinched to five nodes 9 days later. Temperatures in the chambers ranged from a night average of 18°C (SE ± 0.2) to a day average of 27°C (SE ± 0.5). The 24-h mean was 24°C (SE ± 0.2). The experiment was terminated 15 June. Data collected were height of tallest shoot above pot rim, leaf chlorophyll content, stem diameter between third and fourth node of terminal lateral branch, internode length (average length from the third to fifth nodes) of terminal lateral branch, number of leaves on terminal lateral branch, and leaf area. A leaf disk was taken from each of the five most recently expanded leaves from the terminal lateral branch and chlorophyll was extracted and analyzed as already described. The experiment was repeated on 13 June and terminated 22 July.

**Influence of spectral filters on pinched ‘Yellow Mandalay’ chrysanthemums** (Expt. 3). Experimental conditions were the same as for Expt. 2, except ‘Yellow Mandalay’ was used (Yoder Bros.). Temperatures in the chambers ranged from a night average of 19°C (SE ± 0.3) to a day average of 28°C (SE ± 0.5); the 24-h mean was 23°C (SE ± 0.2). Because no differences between air- and water-filled control chambers had been observed in the first two experiments, only water-filled panels served as controls for this experiment. The experiment commenced on 16 June. The plants were pinched 10 days after planting, and the experiment was terminated 5 Aug. Data collection was similar to that in Expt. 2, except internode length was not measured. The experiment was repeated on 4 Aug., terminating on 21 Sept. All plants received natural photoperiods.

Experimental repeats were pooled where no differences occurred between repeats. Data were subjected to analysis of variance (ANOVA), and treatment sums of squares were partitioned among preplanned orthogonal contrasts.

**Results**

**Influence of spectral filters on nonpinched ‘Spears’ chrysanthemums.** Plants grown under CuSO₄ filters were shorter (40%) (Table 2), with shorter internodes (47%) and a higher leaf chlorophyll content (22%) (Table 3) than controls. There were no height, internode, or chlorophyll differences between plants under the red- or blue-dye filters and controls.

There were no differences in stem diameter (data not shown), number of leaves per plant (16.5 to 16.9), and leaf area (280 to 312 cm²) between plants grown under CuSO₄ or dye filters and controls.

**Influence of spectral filters on pinched ‘Spears’ chrysanthemum.** Pinched ‘Spears’ plants grown under CuSO₄ filters were shorter (46%) and had shorter internodes (39%) than the controls (Table 2). There was no difference in internode length between plants grown under blue or red dye and the controls.

<table>
<thead>
<tr>
<th>Filter</th>
<th>Nonpinched</th>
<th>Pinched</th>
<th>Yellow Mandalay, pinched³</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Internode</td>
<td>Internode</td>
<td>pinched³</td>
</tr>
<tr>
<td>Ht (cm)</td>
<td>length (cm)</td>
<td>length (cm)</td>
<td></td>
</tr>
<tr>
<td>Red dye</td>
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<td>1.6</td>
<td>29.4</td>
</tr>
<tr>
<td>Blue dye</td>
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<td>2.0</td>
<td>29.6</td>
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<td>CuSO₄</td>
<td>7.6</td>
<td>0.9</td>
<td>15.5</td>
</tr>
<tr>
<td>Control</td>
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<td></td>
</tr>
<tr>
<td>Control vs. red dye</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
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<tr>
<td>Control vs. blue dye</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
</tr>
<tr>
<td>Control vs. CuSO₄</td>
<td>**</td>
<td>**</td>
<td>**</td>
</tr>
</tbody>
</table>

³Control: data pooled for water- and air-filled filters.
²Control: data for water-filled filters only.
⁴NS, **NS: nonsignificant or significant at P = 0.05 or 0.01, respectively, within columns.

**Table 3. Leaf chlorophyll content of Dendranthema × grandiflorum as influenced by spectral filters.**

<table>
<thead>
<tr>
<th>Filter</th>
<th>Nonpinched</th>
<th>Pinched</th>
<th>Yellow Mandalay, pinched³</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Red dye</td>
<td>33.6</td>
<td>34.3</td>
<td>36.7</td>
</tr>
<tr>
<td>Blue dye</td>
<td>34.0</td>
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<td>39.5</td>
</tr>
<tr>
<td>CuSO₄</td>
<td>42.7</td>
<td>50.5</td>
<td>55.4</td>
</tr>
<tr>
<td>Control</td>
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<td>36.1</td>
<td>39.8</td>
</tr>
<tr>
<td>Contrasts (probability)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control vs. red dye</td>
<td>NS</td>
<td></td>
<td>**</td>
</tr>
<tr>
<td>Control vs. blue dye</td>
<td>NS</td>
<td></td>
<td>NS</td>
</tr>
<tr>
<td>Control vs. CuSO₄</td>
<td>**</td>
<td></td>
<td>**</td>
</tr>
</tbody>
</table>

³Control: data pooled for water- and air-filled filters.
²Control: data for water-filled filters.
⁴NS, **NS: nonsignificant or significant at P = 0.05 or 0.01, respectively, within columns.
Leaf chlorophyll concentration was higher in plants grown under CuSO₄ (39%), less when plants were grown under red dye (5%), and similar when plants were grown under blue dye when contrasted with plants that had no alteration in light quality (Table 3). Stem diameter did not vary between plants grown under CuSO₄ or dye filters and the controls (data not shown).

Plants grown under CuSO₄ had significantly fewer leaves (46%) than the controls (Table 4). Plants grown under dye filters did not differ from the controls. Capitula developed on all CuSO₄-filter-grown plants 10 days before similar structures were visible on plants from other treatments, but they were abnormal. At termination, all lateral branches in CuSO₄-filter treatments had visible capitula, while plants of the other treatments had some vegetative and some reproductive buds (data not shown).

Leaf area of plants grown under CuSO₄ solution or dyes did not differ from that of the controls (Table 4). For all treatments, primary lateral branches developed at all nodes below the pinch.

Influence of spectral filters on pinched ‘Yellow Mandalay’ chrysanthemums. Pinched ‘Yellow Mandalay’ plants were significantly shorter (41%) when grown under the CuSO₄ filter than under control filters. Plants grown under red-dye filters were taller (6%) than the controls, but plants grown under the blue-dye filters were not different from controls (Table 2).

Leaf chlorophyll content was higher (51%) in plants grown under the CuSO₄ filter, lower (8%) in plants under red-dye filters but similar in plants of the blue dye treatments when compared to controls (Table 3).

Leaf count per lateral was inconsistent between experiments. Plants grown under CuSO₄ filters had fewer leaves than control plants in both experiments. Plants grown under blue or red dye were similar to controls in Expt. 1, but had more leaves than controls in Expt. 2 (Table 4). Capitula appeared 3 to 6 days earlier on plants grown under CuSO₄ filters than in plants of other treatments. At termination of the first replication, only plants grown under CuSO₄ filters had capitula visible on all lateral branches. At termination of the second replication, all lateral branches of all treatments had capitula.

Stem diameters and leaf areas of plants grown under CuSO₄ solution or dye-grown plants were similar to those of control plants. Primary lateral branches developed at all nodes below the pinch for all treatments.

Discussion

Without exception, plants grown under the CuSO₄ filters (increased R : FR, increased P₉ : P₁₀) were shorter (Table 2) and had a higher leaf chlorophyll content (Table 3) than controls. These results are consistent with those of Mortensen and Stromme (1987), who determined that ‘Refour’ chrysanthemum, when grown nonpinched under CuSO₄ were shorter and darker green than plants grown under other filters or natural light. At the beginning of our experiments, we were concerned that our chambers had one light-transmitting side and three reflective walls. Reflected light influences plant morphology, particularly if the surfaces reflect radiation selectively (Ballare et al., 1990; Kasperbauer, 1988). The reflective surfaces of our chambers were not selective, and the consistency of our results with those of Mortensen and Stromme’s (whose chambers had four light-transmitting sides) (Mortensen et al., 1987) suggests that chamber construction did not influence plant morphology.

Reduction in plant height was consistent with a reduction in internode length in Expts. 1 and 2 (Table 2). Although internodes were not measured in Expt. 3, calculating the average internode length using plant height and the number of leaves (nodes) per stem indicated that internodes were 25% to 35% shorter for plants grown in CuSO₄-filtered chambers.

Plants that are grown in deficient photosynthetic light are tall, thin, and often etiolated (Cosgrove, 1986). The CuSO₄ filters transmitted light with less intensity at the chlorophyll absorption maxima compared to control filters because some R light was not transmitted. However, even in a less photosynthetically active light environment (McCre, 1972), plants grown in the CuSO₄-altered light environment were shorter and darker green than plants from the control filter chambers.

The reduction of height for pinched plants grown under CuSO₄ filters was partially the result of fewer nodes developing, as indicated by reduced leaf count. Pinched plants grown under CuSO₄ filters developed capitula sooner than other treatments, which caused cessation of node production. However, the capitula were abnormal and it is unlikely that they would have developed into normal flowers.

Chrysanthemums are quantitative short-day plants and can be kept vegetative by long photoperiods or by interrupting long dark periods with light; however, vegetative buds eventually become reproductive (Cockshull, 1975). Flower buds induced under long days do not develop into normal flowers (Laurie et al., 1968). Interrupting the dark period with R light (increased R : FR, increased P₉ : P₁₀) is most effective in preventing floral initiation (Borthwick and Cathey, 1962).

Our study suggests that, during long photoperiods, increased R : FR promotes the change from vegetative to reproductive growth. However, a short photoperiod may be necessary for proper floral development.

Cultivars may also respond differently to increased R : FR. ‘Spears’ grown with increased R : FR developed capitula earlier and with fewer nodes than ‘Yellow Mandalay’ under a similar environment (Table 4). Response to photoperiodic flowering stimulus varies among cultivars (Langton, 1977). ‘Yellow Mandalay’ requires more short days to initiate flowers than ‘Spears’. The cultivar difference may explain why ‘Yellow Mandalay’ grown under CuSO₄ filters produced more nodes before floral initiation than ‘Spears’ even though the natural photoperiod was shorter and temperatures lower for ‘Yellow Mandalay’ than ‘Spears’.

Decreased R : FR or a decrease in B light usually did not appear to affect reproduction. However, development of more nodes on plants grown under the dye filters in the second replication of pinched ‘Yellow Mandalay’ may indicate that reduced R : FR or decreased B light retards floral initiation under

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### Table 4. Leaf count and area on terminal branches of pinched Den-dranthema ×grandiflourum as influenced by spectral filters.

<table>
<thead>
<tr>
<th>Filter</th>
<th>Spears*</th>
<th>Yellow Mandalay*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>No. leaves</td>
<td>Leaf area (cm²)</td>
</tr>
<tr>
<td></td>
<td>Expt. 1</td>
<td>Expt. 2</td>
</tr>
<tr>
<td>Red dye</td>
<td>18.4</td>
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</tr>
<tr>
<td>Blue dye</td>
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</tr>
<tr>
<td>CuSO₄</td>
<td>9.8</td>
<td>1198</td>
</tr>
<tr>
<td>Control</td>
<td>18.0</td>
<td>1304</td>
</tr>
</tbody>
</table>

Contrasts (probability):
- Control vs. red dye:
  - Control vs. blue dye:
  - Control vs. CuSO₄:

*Control: data pooled for water- and air-filled filters.
*Control: data for water-filled filters.
**Nonsignificant or significant at P = 0.05 or 0.01, respectively, within columns.
decreasing photoperiods (the replication ran from 5 Aug. to 21 Sept.).

The increase in chlorophyll in plants with an increase in R : FR compares favorably with another study where Chenopodium alba grown under an artificial light regime of increased R : FR had a higher chlorophyll content than plants grown under a decreased R : FR light regime (Holmes and Smith, 1977).

Plants grown under blue-dye filters did not differ in chlorophyll content from controls. However, plants grown under red-dye filters contained less chlorophyll than the controls. In another study (Leong et al., 1985), Asplenium australasicum grown under R light contained less chlorophyll than those grown under B or white light (R : FR unknown for regimes). Both studies indicate that B light promotes chlorophyll development.

Leaf area of plants grown under CuSO4 filters did not differ from that of controls, although in the experiments with pinched plants, there were fewer leaves per stem. However, we noted that leaves on secondary lateral branches were more expanded on plants filtered by CuSO4 than on plants from the other treatments; these additional leaves would increase the total leaf area. The lack of differences in the experiment with non-pinched plants varies from the findings of Mortensen and Strømme (1987), who observed a reduction in leaf area for plants grown under CuSO4 filters. However, the durations of their experiments were longer and additional time may have allowed leaf area differences to develop. The PPF between the two sites may be different. Mortensen and Strømme reported cumulative PPF over a 30-day period. Therefore, a precise comparison of PPF for the sites is not possible.

The red dye treatment created a blue-deficient environment. However, the plants did not differ in appearance from control plants (reduced chlorophyll levels were not visually apparent). This result differs from those of Dritz and Sager (1990), who reported that a B-light-deficient environment resulted in shade-type plants, i.e., plants were taller and weighed less than those grown under broad-spectrum (daylight fluorescence) lights. However, Dritz and Sager used a narrow-band light source (low-pressure sodium lamps $\lambda_{\text{max}} = 589$ nm). The transmittance of a broad range of wavelengths above 500 nm in our study may have overcome the deficiency in B light. Likewise, plants under the blue-dye filter did not differ from controls. A filter that more closely mimics an understory environment (both R and B light removed) is necessary to induce visible shade-type morphology when broad bands of radiation are present. Reduced chlorophyll levels in red-dye-filtered plants indicate that anatomical changes may be occurring in red- and blue-dye-filtered plants.

We have demonstrated that growth of Dendranthema $\times$ grandiflorum can be manipulated by altering the light environment. Compact, dark-green plants desirable for pot-plant production can be produced under a filter that increases R : FR and increases $P_r : P_{\text{tot}}$. However, unexpected reproductive and morphological responses indicate the need for further study. The filters we used did not allow us to investigate the effects of an environment deficient in both B and FR. Many photomorphogenic responses have been shown to depend on the presence of B light (Cosgrove, 1986).

**Literature Cited**


